



State-of-the-art review of secondary pulmonary infections in patients with COVID-19 pneumonia

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Abstract

Background The incidence of secondary pulmonary infections is not well described in hospitalized COVID-19 patients. Understanding the incidence of secondary pulmonary infections and the associated bacterial and fungal microorganisms identified can improve patient outcomes.

Objective This narrative review aims to determine the incidence of secondary bacterial and fungal pulmonary infections in hospitalized COVID-19 patients, and describe the bacterial and fungal microorganisms identified.

Method We perform a literature search and select articles with confirmed diagnoses of secondary bacterial and fungal pulmonary infections that occur 48 h after admission, using respiratory tract cultures in hospitalized adult COVID-19 patients. We exclude articles involving co-infections defined as infections diagnosed at the time of admission by non-SARS-CoV-2 viruses, bacteria, and fungal microorganisms.

Results The incidence of secondary pulmonary infections is low at 16% (4.8–42.8%) for bacterial infections and lower for fungal infections at 6.3% (0.9–33.3%) in hospitalized COVID-19 patients. Secondary pulmonary infections are predominantly seen in critically ill hospitalized COVID-19 patients. The most common bacterial microorganisms identified in the respiratory tract cultures are *Pseudomonas aeruginosa*, *Klebsiella* species, *Staphylococcus aureus*, *Escherichia coli*, and *Stenotrophomonas maltophilia*. *Aspergillus fumigatus* is the most common microorganism identified to cause secondary fungal pulmonary infections. Other rare opportunistic infection reported such as PJP is mostly confined to small case series and case reports. The overall time to diagnose secondary bacterial and fungal pulmonary infections is 10 days (2–21 days) from initial hospitalization and 9 days (4–18 days) after ICU admission. The use of antibiotics is high at 60–100% involving the studies included in our review.

Conclusion The widespread use of empirical antibiotics during the current pandemic may contribute to the development of multidrug-resistant microorganisms, and antimicrobial stewardship programs are required for minimizing and de-escalating antibiotics. Due to the variation in definition across most studies, a large, well-designed study is required to determine the incidence, risk factors, and outcomes of secondary pulmonary infections in hospitalized COVID-19 patients.

Keywords Severe acute respiratory syndrome coronavirus 2 · SARS-CoV-2 · Coronavirus disease 2019 · COVID-19 · Secondary infection · Superimposed infection · Superinfection · Bacterial infection · Fungal infection

Introduction

Since coronavirus disease 2019 (COVID-19) was first recognized in December 2019, it has resulted in the ongoing worldwide pandemic. COVID-19 is caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), an enveloped RNA beta-coronavirus. SARS-CoV-2 shares a similar genetic identity with severe acute respiratory syndrome coronavirus (SARS-CoV) and belongs to the sarbecovirus subgenus of the Coronaviridae family [1]. COVID-19 primarily presents as a respiratory tract infection with

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symptoms varying from mild flu-like illness to acute respiratory distress syndrome (ARDS) [2, 3]. Viral-related respiratory infections belonging to the same family of coronaviruses such as SARS-CoV and Middle East respiratory syndrome coronavirus (MERS-CoV) have been reported to be associated with secondary bacterial and fungal infections [4–7]. However, secondary pulmonary infections in COVID-19 patients are not well described and raised an important knowledge gap. Furthermore, other infectious and non-infectious complications have been described in hospitalized COVID-19 patients strongly associated with underlying COVID-19 infection such as pneumothorax, myocarditis, and even device-related secondary infections (e.g., central venous catheter, foley catheter). [8–10]. The aim of this review is to explore the incidence of secondary bacterial and fungal pulmonary infections in hospitalized patients with COVID-19 infection. We also discuss the bacterial and fungal microorganisms identified, the time to diagnose secondary pulmonary infections, and the frequency of antibiotic use in hospitalized COVID-19 patients with suspected or confirmed secondary pulmonary infections. There is a lack of data in terms of well-defined risk factors or predictors, and associated outcomes of secondary pulmonary infections in hospitalized patients with COVID-19 infection and, therefore, will not be a major focus of this review.

Method

A literature search was performed through MEDLINE, Pubmed, and Google Scholar using keywords of “coronavirus disease 2019 (COVID-19),” “severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2),” “secondary infection,” “superimposed infection,” “superinfection,” “bacterial infection,” “fungal infection,” “bacterial pneumonia,” “fungal pneumonia,” “bacteremia,” “fungemia,” “hospital-acquired pneumonia (HAP),” and “ventilator-associated pneumonia (VAP)” from January 1st, 2020 to December 31st, 2020. Our selection criteria comprised of articles with confirmed diagnoses of secondary bacterial and fungal pulmonary infections (defined as new microorganisms identified 48 h after admission) using respiratory tracts with corresponding blood cultures for similar microorganisms thought to be respiratory in origin in hospitalized adult COVID-19 patients. Respiratory tract cultures were defined as cultures obtained from sputum, endotracheal aspirates, and bronchoalveolar lavage (BAL). We also included articles in which the diagnoses of secondary pulmonary infections were suspected based on the description of cultures obtained that were respiratory in nature or microorganisms that are recognized to be respiratory in origin. Articles published in the English language were selected, and any cited references were reviewed to identify relevant literature in

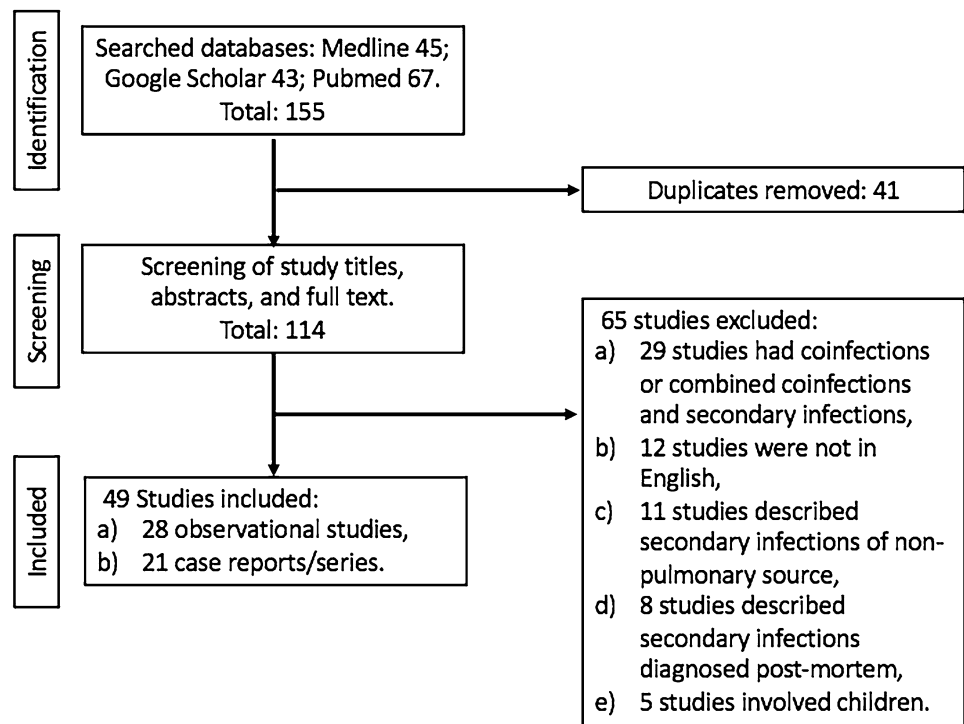
the English language that comprised of observational studies, case reports, and series that met our selection criteria that described secondary pulmonary infections in hospitalized COVID-19 patients. We excluded articles involving COVID-19 infections in children and pregnant women; non-hospitalized COVID-19 patients; patients with pulmonary co-infections (defined as infections diagnosed at the time of admission) by non-SARS-CoV-2 viruses, bacteria, and fungal microorganisms; secondary pulmonary infections from microorganisms that were known to be colonizers such as candida; studies that included both secondary infections and co-infections from a non-pulmonary source in hospitalized COVID-19 patients; and the diagnosis of secondary pulmonary infections made during the post-mortem examination of deceased COVID-19 patients. We screened 114 studies and included 49 studies that described secondary pulmonary infections in hospitalized adult COVID-19 patients that met our criteria (Fig. 1). Of the 12 studies published in the non-English language (defined as non-English language articles and had no English translation versions/options) that described secondary pulmonary infections in hospitalized COVID-19 patients, 6 were published in Mandarin, 4 were published in Spanish, and the remaining 2 were published in French. The diagnosis of COVID-19 was made by reverse transcriptase-polymerase chain reaction (RT-PCR) in all cases from respiratory tract specimens that include nasal and pharyngeal swabs, sputum, endotracheal aspirates, and bronchoalveolar lavage (BAL).

Results

Incidence of secondary pulmonary infections

Among the 49 studies identified (Table 1), 28 (57%) studies were observational studies done on hospitalized COVID-19 patients; the remainder, 21 (43%), were small case series and case reports. Of the 28 observational studies, 78.6% were retrospective and 21.4% were prospective in nature. The majority of observational studies originated from China in 25% (7/28) of cases followed by 17.9% (5/28) in Spain, 14.3% (4/28) in France, 7.1% in Netherlands and USA, respectively, and the remainder in Belgium, Denmark, England, Germany, Italy, Mexico, Pakistan, and Switzerland. A total of 5,047 hospitalized patients with COVID-19-related pneumonia were identified in the 28 observational studies included in our review (Table 1). The incidence of secondary bacterial pulmonary infections in hospitalized COVID-19 patients reported was 16% (580/3,633) and ranged between 4.8–42.8% in 14 observational studies, whereas the incidence of secondary fungal infections in hospitalized COVID-19 patients was 6.3% (171/2,703) and ranged between 0.9 and 33.3% according to 18 observational studies

Fig. 1 Flowchart for Studies Selected in Review of Hospitalized COVID-19 Patients With Secondary Pulmonary Infections



(Table 1). The majority of hospitalized COVID-19 patients who developed secondary bacterial and fungal infections were critically ill where they required ICU admission and invasive mechanical ventilation (IMV). Studies by Chang et al. , Rouze et al. , and Torrego et al. were the only observational studies that examined the incidence of secondary bacterial pulmonary infection based solely on BAL findings in COVID-19 patients requiring IMV with moderate to severe ARDS [11–13]. However, for examining the incidence of secondary fungal pulmonary infections, respiratory cultures obtained from BAL were only used in observational studies by Bartoletti et al. , Rutsaert et al. , and Van Biesen et al. [14–16].

Microbiology of secondary pulmonary infections

Out of the 28 observational studies, 14.3% (4/28) of studies had no descriptions of the specific bacterial or fungal microorganisms identified (Table 2). The most common bacterial microorganisms identified in the respiratory tract cultures among the nine observational studies (Table 2) that reported the type and frequency of secondary bacterial infection were 21.1% (75/355) *Pseudomonas aeruginosa*, 17.2% (61/355) *Klebsiella* species, 13.5% (48/355) *Staphylococcus aureus*, 10.4% (37/355) *Escherichia coli*, and 3.1% (11/355) *Stenotrophomonas maltophilia*. However, the fungal microorganisms identified in 18 observational studies included in our review were predominantly *Aspergillus* species in which *Aspergillus fumigatus* was most frequently isolated in all

studies. Other less common *Aspergillus* species identified were *Aspergillus flavus*, *Aspergillus calidoustus*, *Aspergillus citrinoterreus*, *Aspergillus niger*, *Aspergillus terreus*, and *Aspergillus versicolor*. One observational study by Fekkar et al. reported an uncommon finding of *Mucor* species and *Fusarium proliferatum* in respiratory tract cultures of critically ill COVID-19 patients [17]. Other rare opportunistic fungal infections such as *Pneumocystis jirovecii* (PJP) had been observed in four case reports/series included in our review [18–21].

The time to diagnosis of secondary pulmonary infections and use of antibiotics

The average time taken to diagnose secondary bacterial and fungal pulmonary infections from hospital and ICU admission among the 18 observational studies described was 10 days (ranged 2–21 days) and 9 days (ranged 4–18 days), respectively (Table 2). The reported use of empirical antibiotics was 60–100% during the current pandemic between 11 observational studies (Table 2). Furthermore, although specific data on antibiotic resistance patterns lacked in the majority of observational studies included in our review, limited observational studies had reported of detection of multidrug-resistant (MDR) microorganisms such as extended-spectrum beta-lactamase (ESBL) *Klebsiella pneumoniae*, ESBL *Escherichia coli*, MDR *Pseudomonas aeruginosa*, carbapenem-resistant *Klebsiella pneumoniae*, and Methicillin-resistant *Staphylococcus aureus* (MRSA)

Table 1 Characteristics of Secondary Pulmonary Infections Studies in Hospitalized COVID-19 Patients With Incidence of Secondary Bacterial and Fungal Infections

| Study | Region | Study Type | Total COVID-19 Patients (N) | Age | ICU (%) | IMV (%) | Hospital Mortality* (%) | Secondary bacterial infections (%) | Secondary fungal infections (%) |
|-----------------------------|-------------|-----------------------------|-----------------------------|--------------|---------|---------|-------------------------|------------------------------------|---------------------------------|
| Observational Studies | | | | | | | | | |
| Alanio et al. [43] | France | Retrospective, ICU | 27 | 63 | 100 | 100 | 44.4 | NR | 33.3 |
| Barrasa et al. [25] | Spain | Retrospective, ICU | 48 | 63 (mean) | 100 | 93.7 | 35.4 | 12.5 | NR |
| Bartoletti et al. [14] | Italy | Prospective, ICU | 108 | 63* (median) | 100 | 100 | 44.0 | NR | 27.7 |
| Chang et al. [12] | USA | Retrospective, ICU | 412 | 62 (median) | 100 | 100 | NR | 15.5 | NR |
| Du et al. [56] | China | Prospective, Hospital | 179 | 58 (median) | NR | NR | 11.7 | 5.6 | NR |
| Dupont et al. [69] | France | Prospective, ICU | 106 | 69* (median) | 100 | 100 | 35.3 | NR | 17.9 |
| Fekkar et al. [17] | France | Retrospective, ICU | 145 | 55 (median) | 100 | 100 | 57.1 | NR | 4.8 |
| Feng et al. [32] | China | Retrospective, Hospital/ICU | 410 | 53 (median) | 14.8 | 8.0 | 8.0 | 9.0 | NR |
| Fu et al. [23] | China | Retrospective, Hospital/ICU | 101 | 69* (mean) | 35.6 | 100 | NR | 5.0 | 0.9 |
| Gangneux et al. [70] | France | Prospective, ICU | 45 | 70* (mean) | 100 | 100 | 28.6 | NR | 15.6 |
| Garcia-Vidal et al. [22] | Spain | Retrospective, Hospital/ICU | 989 | 62 (median) | 11.9 | NR | 9.8 | 4.3 | 0.7 |
| Helleberg et al. [71] | Denmark | Retrospective, ICU | 25 | 58 | 100 | 100 | 100 | NR | 8.0 |
| Huang et al. [30] | China | Retrospective, Hospital/ICU | 41 | 49 (median) | 46.0 | 5.0 | 15.0 | 10.0 | NR |
| Karmen-Tuohy et al. [58] | USA | Retrospective, Hospital | 63 | 60 (median) | 20.6 | 15.8 | 25.3 | 6.3 | NR |
| Lamoth et al. [72] | Switzerland | Retrospective, ICU | 118 | 62* (mean) | 100 | 100 | 33.3 | NR | 3.8 |
| Machado et al. [35] | Spain | Prospective, ICU | 239 | 63* (mean) | 100 | 100 | 100 | NR | 2.5 |
| Nasir et al. [26] | Pakistan | Retrospective, Hospital/ICU | 147 | 71 (median) | 15.6 | 40 | 60.0 | 4.8 | 6.1 |
| Roman-Montes et al. [73] | Mexico | Retrospective, ICU | 144 | 48* (mean) | 100 | NR | 57.1 | NR | 9.7 |
| Rouze et al. [13] | Germany | Retrospective, ICU | 568 | 64 (mean) | 100 | 100 | NR | 36.1 | NR |
| Rutsaert et al. [15] | Belgium | Retrospective, ICU | 34 | 66 | 100 | 100 | 58.8 | NR | 20.5 |
| Segrelles-Calvo et al. [74] | Spain | Retrospective, ICU | 215 | 60 (median) | 100* | 100* | 71.4 | NR | 3.3 |

Table 1 (continued)

| Study | Region | Study Type | Total COVID-19 Patients (N) | Age | ICU (%) | IMV (%) | Hospital Mortality* (%) | Secondary bacterial infections (%) | Secondary fungal infections (%) |
|---------------------------|-------------|-----------------------------|-----------------------------|-------------|----------|----------|-------------------------|------------------------------------|---------------------------------|
| Torrego et al. [11] | Spain | Retrospective, ICU | 93 | NR | 100 | NR | NR | 19.3 | NR |
| Van Arkel et al. [75] | Netherlands | Retrospective, ICU | 31 | 64 | 100 | 83.9 | 67.7 | NR | 19.4 |
| Van Biesen et al. [16] | Netherlands | Retrospective, ICU | 42 | 68 (mean) | 100 | 100 | 22.2 | NR | 21.4 |
| Wang et al. [57] | China | Retrospective, Hospital/ICU | 339 | 71 (median) | NR | 23.6 | 19.1 | 42.8 | NR |
| White et al. [34] | England | Prospective, ICU | 135 | 57 (median) | 100 | 72.0 | 52.0 | NR | 18.5 |
| Yang et al. [24] | China | Retrospective, ICU | 52 | 60 (mean) | 100 | 71.0 | 61.5 | 7.7 | 3.8 |
| Zhou et al. [31] | China | Retrospective, Hospital/ICU | 191 | 56 (median) | 26.0 | 17.0 | 28.2 | 15.0 | NR |
| Case Series/Reports | | | | Age* (mean) | ICU* (%) | IMV* (%) | Death* (%) | Secondary bacterial infections (%) | Secondary fungal infections (%) |
| Abdalla et al. [76] | Qatar | Case Series, ICU | 2 | 66 | 100 | 100 | 100 | NR | 100 |
| Blanco et al. [19] | Spain | Case Series, Hospital | 5 | 38 | 40.0 | 20.0 | 0.0 | NR | 20.0 |
| Falces-Romero et al. [77] | Spain | Case Series, Hospital | 10 | 70 | 70.0 | 70.0 | 70.0 | NR | 100 |
| Koehler et al. [78] | Germany | Case Series, ICU | 19 | 63 | 100 | 100 | 57.8 | NR | 26.3 |
| Lahmer et al. [79] | Germany | Case Series, ICU | 2 | 75 | 100 | 100 | 100 | NR | 100 |
| Lescure et al. [80] | France | Case Series, Hospital | 5 | 47 | 60.0 | 40.0 | 40.0 | 20.0 | 20.0 |
| Sharifipour et al. [81] | Iran | Case Series, ICU | 19 | 67 | 100 | 100 | 94.7 | 100 | NR |
| Antinori et al. [82] | Italy | Case Report, ICU | 1 | 56 | 100 | 100 | 100 | NR | 100 |
| Blaize et al. [83] | France | Case Report, ICU | 1 | 74 | 100 | 100 | 100 | NR | NR |
| Duployez et al. [84] | France | Case Report, ICU | 1 | 30 s | 100 | 100 | 100 | NR | NR |
| Fernandez et al. [85] | Argentina | Case Report, ICU | 1 | 85 | 100 | 100 | 100 | 100 | 100 |
| Ghelfenstein et al. [86] | France | Case Report, ICU | 1 | 56 | 100 | 100 | 100 | NR | 100 |
| Kelly et al. [21] | England | Case Report, ICU | 1 | 50 s | 100 | 100 | 100 | NR | 100 |
| Mang et al. [20] | Germany | Case Report, ICU | 1 | 52 | 100 | 100 | 0.0 | NR | 100 |
| Meijer et al. [87] | Netherlands | Case Report, ICU | 1 | 74 | 100 | 100 | 100 | NR | 100 |

Table 1 (continued)

| Case Series/Reports | | | | Age* (mean) | ICU* (%) | IMV* (%) | Death* (%) | Secondary bacterial infections (%) | Secondary fungal infections (%) |
|---------------------|-----------|-----------------------|---|-------------|----------|----------|------------|------------------------------------|---------------------------------|
| Menon et al. [18] | USA | Case Report, ICU | 1 | 83 | 100 | 100 | 0.0 | NR | NR |
| Mohamed et al. [47] | Ireland | Case Report, ICU | 1 | 66 | 100 | 100 | 100 | 100 | 100 |
| Nasri et al. [88] | Iran | Case Report, ICU | 1 | 42 | 100 | 100 | 100 | NR | 100 |
| Prattes et al. [89] | Austria | Case Report, ICU | 1 | 70 | 100 | 100 | 100 | NR | 100 |
| Schein et al. [90] | France | Case Report, Hospital | 1 | 87 | 0 | 0 | 100 | NR | 100 |
| Sharma et al. [91] | Australia | Case Report, ICU | 1 | 66 | 100 | 100 | 0 | NR | 100 |

*Among those with secondary infections

ICU = intensive care unit, IMV = invasive mechanical ventilation, NR = not reported

from the respiratory tract and blood cultures in critically ill COVID-19 patients [22–26].

Discussion

In hospitalized COVID-19 patients, the incidence of secondary pulmonary infections was low at 16% (4.8–42.8%) for bacterial infections and lower for fungal infections with an incidence of 6.3% (0.9–33.3%). However, the frequency of empirical antibiotic therapy was high at 60–100% among several observational studies included. The most common bacterial microorganisms identified in the respiratory tract cultures were 21.1% *Pseudomonas aeruginosa*, 17.2% *Klebsiella* species, 13.5% *Staphylococcus aureus*, 10.4% *Escherichia coli*, and 3.1% *Stenotrophomonas maltophilia*. *Aspergillus fumigatus* was the most common fungal microorganism identified to cause secondary pulmonary infections. Other rare opportunistic infection such as PJP was mostly confined to small case series and case reports. The overall time to diagnose secondary bacterial and fungal pulmonary infections was 10 days (2–21 days) and 9 days (4–18 days), respectively, from the time of hospital and ICU admission.

In contrast, the incidence of secondary bacterial pulmonary infections during the 2009 Influenza A pandemic is up to 7% in critically ill patients [27]. However, for secondary fungal pulmonary infections, the incidence is as high as 14% in critically ill patients with seasonal influenza [28], 29. A retrospective study by Rouze et al. reported that secondary bacterial pulmonary infections were 1.6 times more likely to occur in critically ill COVID-19 patients compared to influenza patients. Four observational studies did not report any specific type of

microorganism identified [3, 30–32]. In these studies, secondary pulmonary infections were minor secondary outcomes identified while assessing the many characteristics, risk factors, and outcomes of hospitalized COVID-19 patients. Furthermore, although 18 observational studies described secondary fungal pulmonary infections predominantly *Aspergillus fumigatus*, there was an absence of a standardized definition with the heterogeneity of diagnostic criteria used to differentiate between true infection versus colonization [33–35]. The microorganisms identified to cause secondary bacterial pulmonary infections in hospitalized COVID-19 patients are similar to microorganisms isolated during seasonal/pandemic influenza and even during the 2003 SARS outbreak [13, 36, 37]. The identification of gram-negative microorganisms in hospitalized COVID-19 patients is consistent with the type of pathogens commonly associated with hospital-acquired pneumonia involving *Pseudomonas aeruginosa*, *Klebsiella* species, *Stenotrophomonas maltophilia*, and *Acinetobacter baumannii* that does not necessarily suggest a specific preference for gram-negative infections in COVID-19 [38–41]. The time taken for the diagnosis of secondary pulmonary infections is highly variable between 2 and 21 days from hospital admission and 4–18 days from ICU admission according to the 18 observational studies included in our review (Table 2). This differs in contrast to secondary bacterial infections that are diagnosed earlier in patients with influenza infection, which are 3–6 days from the initial presentation [36, 42]. For secondary invasive pulmonary aspergillosis in influenza patients, the median time to diagnosis is between 5 and 10 days after ICU admission [15, 29]. Although all observational studies included described respiratory tract cultures obtained more than 48 h after

Table 2 Studies in Hospitalized COVID-19 Patients With Types of Microorganisms Reported, Rate of Antibiotic Use, and Time to Diagnosis of Secondary Pulmonary Infections

| Study | Top Five Bacterial Microorganisms From Respiratory Tract Culture | Fungal Microorganisms From Respiratory Tract Culture | Empirical Antibiotic Therapy (%) | Median Day to Secondary Pulmonary Infection Diagnosis From Admission |
|------------------------------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------------------------|----------------------------------|----------------------------------------------------------------------|
| Observational Studies | | | | |
| Alantio et al. [43] | NR | 100% <i>Aspergillus fumigatus</i> | NR | NR |
| Barrasa et al. [25] | 50% <i>Pseudomonas aeruginosa</i> , 16% <i>Enterococcus faecium</i> , 16% <i>Haemophilus influenzae</i> , 16% Methicillin-Resistant <i>Staphylococcus aureus</i> (MRSA) | NR | 87.5 | NR |
| Bartoletti et al. [14] | NR | 78.9% <i>Aspergillus fumigatus</i> , 15.8% <i>Aspergillus niger</i> , 5.3% <i>Aspergillus flavus</i> | NR | 4 (ICU) |
| Chang et al. [12] | 36% <i>Klebsiella</i> spp., 23% MSSA, 11% <i>Escherichia coli</i> , 11% <i>Serratia</i> spp., 11% <i>Stenotrophomonas</i> spp. | NR | NR | NR |
| Du et al. [56] | <i>Acinetobacter baumannii</i> , <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> , <i>Staphylococcus</i> species** | NR | NR | NR |
| Dupont et al. [69] | NR | 87.5% <i>Aspergillus fumigatus</i> , 6.3% <i>Aspergillus flavus</i> , 6.3% <i>Aspergillus calidoustus</i> | NR | 11 (ICU) |
| Fekkar et al. [17] | NR | 66.7% <i>Aspergillus fumigatus</i> , 22.2% <i>Mucor</i> spp., 11.1% <i>Fusarium proliferatum</i> | NR | 18 (ICU) |
| Feng et al. [32] | NR | NR | 67.1 | NR |
| Fu et al. [23] | 29% <i>Pseudomonas aeruginosa</i> , 29% <i>Burkholderia cepacia</i> , 14% <i>Escherichia coli</i> , 14% Extended-spectrum beta-lactamase (ESBL) <i>Klebsiella pneumoniae</i> , 14% <i>Stenotrophomonas maltophilia</i> | 100% <i>Aspergillus fumigatus</i> | NR | 12 |
| Gangneux et al. [70] | NR | 100% <i>Aspergillus fumigatus</i> | NR | NR |
| Garcia-Vidal et al. [22] | 33% multidrug-resistant (MDR) <i>Pseudomonas aeruginosa</i> , 25% ESBL <i>Escherichia coli</i> , 21% ESBL <i>Klebsiella pneumoniae</i> , 21% <i>Staphylococcus aureus</i> | 100% <i>Aspergillus fumigatus</i> | 61.9 | 11 |
| Helleberg et al. [71] | NR | 100% <i>Aspergillus fumigatus</i> | NR | 3 (ICU) |
| Huang et al. [30] | NR | NR | 100 | >2 |
| Karmen-Tuohy et al. [58] | 29% <i>Stenotrophomonas maltophilia</i> , 29% <i>Pseudomonas aeruginosa</i> , 14% <i>Escherichia coli</i> , 14% <i>Klebsiella pneumoniae</i> , 14% <i>Staphylococcus aureus</i> | NR | NR | >6 |
| Lamoth et al. [72] | NR | 100% <i>Aspergillus fumigatus</i> | 100 | 6 (ICU) |
| Machado et al. [35] | NR | 66.6% <i>Aspergillus fumigatus</i> , 16.7% <i>Aspergillus citrinotereus</i> , 16.7% <i>Aspergillus lentulus</i> | 100 | 15 (ICU) |

Table 2 (continued)

| Study | Top Five Bacterial Microorganisms From Respiratory Tract Culture | Fungal Microorganisms From Respiratory Tract Culture | Empirical Antibiotic Therapy (%) | Median Day to Secondary Pulmonary Infection Diagnosis From Admission |
|-----------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------------------------------------------------|------------------------------------|----------------------------------------------------------------------|
| Nasir et al. [26] | 25% <i>Pseudomonas aeruginosa</i> , 25% <i>Acinetobacter</i> spp., 25% <i>Stenotrophomonas maltophilia</i> , 12.5% <i>Klebsiella pneumoniae</i> , 12.5% MRSA | 60% <i>Aspergillus flavus</i> , 20% <i>Aspergillus fumigatus</i> , 20% <i>Aspergillus terreus</i> | 77.7 | 7 |
| Roman-Montes et al. [73] | NR | 54.5% <i>Aspergillus fumigatus</i> , 27.3% <i>Aspergillus</i> spp., 9.1% <i>Aspergillus flavus</i> , 9.1% <i>Aspergillus niger</i> | NR | 9 |
| Rouze et al. [13] | 22.3% <i>Pseudomonas aeruginosa</i> , 18.8% <i>Enterobacter</i> spp., 11.5% <i>Klebsiella</i> spp., 9.4% MSSA, 8.4% <i>Escherichia coli</i> | NR | 93.2 | NR |
| Rutsaert et al. [15] | NR | 83% <i>Aspergillus fumigatus</i> , 17% <i>Aspergillus Flavus</i> | NR | 6 |
| Segrelles-Calvo et al. [74] | NR | 42.8% <i>Aspergillus fumigatus</i> , 28.6% <i>Aspergillus flavus</i> , 28.6% <i>Aspergillus niger</i> | NR | NR |
| Torrego et al. [11] | 39% <i>Pseudomonas aeruginosa</i> , 11% <i>Enterobacter cloacae</i> , 11% <i>Enterococcus faecalis</i> , 11% <i>Klebsiella aerogenes</i> , 11% <i>Staphylococcus aureus</i> | NR | NR | <21 |
| Van Arkel et al. [75] | NR | 100% <i>Aspergillus fumigatus</i> | NR | 12 |
| Van Biesen et al. [16] | NR | 71.4% <i>Aspergillus fumigatus</i> , 14.3% <i>Aspergillus flavus</i> , 14.3% <i>Aspergillus terreus</i> | NR | 5 (ICU) |
| Wang et al. [57] | NR | NR | NR | NR |
| White et al. [34] | NR | 91.6% <i>Aspergillus fumigatus</i> , 8.4% <i>Aspergillus versicolor</i> | 98.0 | 8 (ICU) |
| Yang et al. [24] | 25% ESBK <i>Klebsiella pneumoniae</i> , 25% <i>Carbapenem-resistant Klebsiella pneumoniae</i> , 25% MDR <i>Pseudomonas aeruginosa</i> , 25% <i>Serratia marcescens</i> | 50% <i>Aspergillus fumigatus</i> , 50% <i>Aspergillus flavus</i> | 94.2 | NR |
| Zhou et al. [31] | NR | NR | 94.8 | 17 |
| Case Series/Reports | Top Five Bacterial Microorganisms From Respiratory Tract Culture | Fungal Microorganisms From Respiratory Tract Culture | Antibiotic/Antifungal Therapy* (%) | Median Day to Secondary Pulmonary Infection Diagnosis From Admission |
| Abdalla et al. [76] | NR | 50% <i>Aspergillus niger</i> , 50% <i>Aspergillus terreus</i> | 100 | 12 (ICU) |
| Blanco et al. [19] | NR | 100% <i>Pneumocystis jirovecii</i> | 60.0 | 12 |
| Falces-Romero et al. [77] | NR | 87.5% <i>Aspergillus fumigatus</i> , 12.5% <i>A. nidulans</i> | 80.0 | 17.1 |
| Koehler et al. [78] | NR | 100% <i>Aspergillus fumigatus</i> | NR | NR |
| Lahmer et al. [79] | NR | 100% <i>Aspergillus fumigatus</i> | NR | 5.5 (ICU) |
| Lescure et al. [80] | 100% <i>Acinetobacter baumannii</i> | 100% <i>Aspergillus fumigatus</i> | NR | NR |
| Sharifipour et al. [81] | 89.4% <i>Acinetobacter baumannii</i> , 5.3% MSSA, 5.3% MRSA | NR | NR | NR |

Table 2 (continued)

| Case Series/Reports | Top Five Bacterial Microorganisms From Respiratory Tract Culture | Fungal Microorganisms From Respiratory Tract Culture | Antibiotic/Antifungal Therapy* (%) | Median Day to Secondary Pulmonary Infection Diagnosis From Admission |
|--------------------------|--------------------------------------------------------------------------------------------------|------------------------------------------------------|------------------------------------|----------------------------------------------------------------------|
| Antinori et al. [82] | NR | 100% <i>Aspergillus fumigatus</i> | 100 | 9 |
| Blaize et al. [83] | NR | 100% <i>Aspergillus fumigatus</i> | 100 | 4 |
| Duployez et al. [84] | 100% Panton-Valentine Leukocidin (PVL) Methicillin-Sensitive <i>Staphylococcus aureus</i> (MSSA) | N/A | 100 | 5 |
| Fernandez et al. [85] | 100% <i>Enterococcus faecalis</i> , <i>Acinetobacter baumannii</i> | 100% <i>Aspergillus fumigatus</i> | 100 | 10 |
| Ghelfenstein et al. [86] | NR | 100% <i>Aspergillus fumigatus</i> | 100 | 6 |
| Kelly et al. [21] | NR | 100% <i>Pneumocystis jirovecii</i> | 100 | 10 |
| Mang et al. [20] | NR | 100% <i>Pneumocystis jirovecii</i> | 100 | 20 |
| Meijer et al. [87] | NR | 100% <i>Aspergillus fumigatus</i> | 100 | 6 |
| Menon et al. [18] | NR | 100% <i>Pneumocystis jirovecii</i> | NR | NR |
| Mohamed et al. [47] | <i>Klebsiella Varicola</i> | 100% <i>Aspergillus fumigatus</i> | 100 | 7 |
| Nasri et al. [88] | NR | 100% <i>Aspergillus spp.</i> | 100 | 9 |
| Prattes et al. [89] | NR | 100% <i>Aspergillus fumigatus</i> | 100 | 3 (ICU) |
| Schein et al. [90] | NR | 100% <i>Aspergillus spp.</i> | 100 | 16 |
| Sharma et al. [91] | NR | 100% <i>Aspergillus fumigatus</i> | 100 | 16 |

*Among those with secondary infections

***Percentage frequency of bacterial or fungal microorganisms not reported

NR not reported, *Spp.* Species

admission, the variability in time to diagnosis can be due to the inconsistency on when and a lack of information on why surveillance cultures are obtained.

Bronchoscopy may be a useful tool to obtain respiratory tract cultures of sufficient quantity to help diagnose and isolate microorganisms in secondary pulmonary infections while determining the antibiotic sensitivities in hospitalized COVID-19 patients. The routine use of bronchoscopy may even lead to over-diagnosis of secondary pulmonary infections from respiratory tract colonization. According to three observational studies, the incidence of secondary bacterial pulmonary infections was 15% and more when routine bronchoscopy with BAL was performed in critically ill COVID-19 patients requiring IMV [11–13]. Four observational studies reported that the incidence of secondary fungal pulmonary infections was 20% and more in critically ill COVID-19 patients when bronchoscopy with BAL was performed routinely post-intubation, in a serial fashion, or any change in clinical status due to atelectasis, new lung infiltrates on imaging, and thick secretions [14–16, 43]. Chang et al. described that respiratory tract cultures obtained from BAL have a higher positivity rate when compared to endotracheal aspirate and a greater tendency to detect different or second microorganisms as a cause of secondary pulmonary infections [12]. In the study by Torrego et al., the microbiology findings on BAL resulted in a change in antibiotic prescribed in 83% of critically ill COVID-19 patients requiring IMV. However, the bacterial microorganisms identified such as *Pseudomonas aeruginosa*, *Klebsiella* species, *Enterobacter cloacae*, and *Staphylococcus aureus* was similar to bacterial microorganisms in mechanically ventilated non-COVID-19 patients [11]. However, bronchoscopy is often avoided as it is an aerosol-generating procedure that will predispose healthcare workers and patients to a substantial risk of further transmitting COVID-19 infection. The use of bronchoscopy in COVID-19 patients has been recommended when current respiratory samples from sputum and endotracheal aspirates are negative, in which an alternate diagnosis provided by BAL would significantly impact clinical management [44]. Nevertheless, two recent single-center retrospective studies showed no increase in the risk of COVID-19 transmission to healthcare providers when bronchoscopy is routinely performed while adhering to the proper infection control protocol [12, 45].

The current knowledge of the risk factors for secondary pulmonary infections in SARS-CoV-2 is continuously evolving but remains poorly understood. Although it is becoming apparent that secondary pulmonary infections that occur in hospitalized COVID-19 patients can be associated with worse outcomes, it remains unclear if critically ill COVID-19 patients are at a greater likelihood of developing secondary pulmonary infections. COVID-19 infection will trigger innate and adaptive immune responses, including

local immune response, recruitment of macrophages and monocytes, the release of cytokines, and prime adaptive T- and B-cell in an effort to resolve underlying inflammation [46–49]. However, in some cases, a dysfunctional immune response occurs that renders COVID-19 patients vulnerable to secondary pulmonary infections. Lymphocyte count, specifically T-cells, is substantially decreased, whereas inflammatory mediators of interleukins- (IL-)2, IL-6, IL-8, IL-10, tumor necrosis factor-alpha (TNF-a), and interferon-gamma are markedly increased within a week from COVID-19 presentation before recovering to normal levels, two weeks later [30, 50–53]. This dysregulated immune response that is seen to a greater degree in those with severe COVID-19 infections has an immunosuppression stage following the proinflammatory phase characterized by a sustained and substantial reduction in peripheral lymphocyte count [48, 50, 54]. Similar immunological findings have been described in SARS-CoV patients during the 2003 epidemic and H1N1 influenza during the 2009 pandemic [53–55]. This state of lymphocytopenia-induced immunosuppression observed in many hospitalized COVID-19 patients may explain the time taken for secondary pulmonary infection diagnosis seen in studies included in our review [30–32, 56].

Furthermore, in a multi-center study involving 410 COVID-19 patients, secondary pulmonary infections were significantly associated with outcome severity. Critically ill patients had the highest percentage of secondary pulmonary infections (34.5%) compared to severely ill (8.3%) and moderately ill (3.9%) COVID-19 patients [32]. This high rate of secondary pulmonary infections occurs despite a majority of critically ill patients (92.9%) receiving antibiotics compared to 83.3% and 59.4% in the severely ill and moderately ill groups. Five observational studies reported that among critically ill COVID-19 patients, non-survivors/critically ill patients had a greater tendency to suffer from multi-organ dysfunction and develop secondary pulmonary infections despite up to 98% of them received antibiotics [24, 30, 31, 56, 57]. In all these studies, the degree of lymphocytopenia and corticosteroids administration was significantly higher in the critically ill/non-survivor group than in other groups. Furthermore, although the nadir CD4 + T-cell count was less than 200 cells/ 10^6 L in the majority of case reports/series describing PJP among HIV patients co-infected with COVID-19 [19–21], a case report by Menon et al. described a hospitalized COVID-19 patient diagnosed with PJP despite the absence of HIV infection. Though her nadir CD4 + T-cell count was 291 cells/ 10^6 L and she was on chronic oral budesonide for her ulcerative colitis, the improvement with trimethoprim-sulfamethoxazole supported the diagnosis of secondary PJP infection [18]. On the contrary, a retrospective study by Karmen-Tuohy et al. reported no increased incidence of secondary bacterial or even PJP pulmonary infections in HIV-positive COVID-19 patients who were

compliant with antiretroviral therapy, regardless of their CD4 + T cell count [58]. There is no single study to the current date, which has formally assessed lymphocytopenia as a risk factor for secondary pulmonary infections. Moreover, corticosteroids are frequently used in COVID-19 patients to prevent and treat cytokine storm and ARDS, which are suspected to be partly caused by dysregulated host immune response [50, 53, 59]. Recent studies assessing the use of corticosteroids in hospitalized COVID-19 patients demonstrated that a short course of corticosteroids over ten days has shown to be beneficial in the setting of hypoxic respiratory failure requiring oxygen therapy and mechanical ventilation requirement [60, 61]. However, previous studies have demonstrated that corticosteroids may inadvertently increase the mortality and secondary infections in influenza patients, and prolong viral shedding and induce lymphocytopenia in SARS-CoV patients by down-regulating the innate and adaptive immune system.[29, 54, 62, 63] Currently, there is no formal study to assess the risk of secondary pulmonary infections associated with corticosteroids administration in COVID-19 patients. However, in our review, although the majority of patients were receiving corticosteroids, the timing of administration, duration, and the dose of corticosteroids were not clearly described.

Differentiating viral from secondary bacterial and fungal pulmonary infections remains a challenge for clinicians. This diagnostic uncertainty has contributed to the overuse of antibiotics in patients with COVID-19 viral illness. Although the incidence of secondary bacterial pulmonary infections in COVID-19 patients is low, the reported use of empirical antibiotics is 60–100% among the observational studies included in our review (Table 2). These findings vastly differed when compared to patients with seasonal/pandemic influenza, in which the reported use of empirical antibiotics was 12–50% [36]. It is essential to consider how the frequent use of empirical antibiotic therapy could affect the prevalence of multidrug-resistant bacteria. The rising number of antibiotic use may predispose COVID-19 patients, especially those who are critically ill, to sepsis from secondary multidrug-resistant bacterial infections. An observational study during the SARS outbreak in 2003 demonstrated that MRSA acquisition identified on screening using nasal swabs drastically increased from 2.2 to 3.5 cases per 100 ICU admissions (pre-SARS and post-SARS period) to 25.3% per 100 ICU admissions (during SARS period), despite extensive infection control precautions [64]. This finding coincides with the increased use of broad-spectrum empiric antibiotics (4th generation cephalosporins, fluoroquinolones, aminoglycosides, and carbapenems) during the SARS period, in which MRSA was responsible for up to 48% of microorganisms isolated in patients with VAP. Furthermore, common bacterial microorganisms identified on post-mortem examination of SARS patients were *Pseudomonas aeruginosa*, *Klebsiella*

species, and *Staphylococcus aureus*, which are known for their high resistance to broad-spectrum antibiotics [65, 66]. In the studies that we reviewed, antibiotic sensitivities of microorganisms and treatment duration were not reported even though MDR microorganisms were observed. Based on the current microbiological data from our review, it remains imperative that empiric antibiotic therapy covers multidrug-resistant microorganisms such as MRSA and ESBL that are associated with a high fatality rate when concerns exist of possible secondary pulmonary infections in critically ill COVID-19 patients [39, 40]. Our review supports the notion of frequently obtaining surveillance cultures (from sputum, endotracheal aspirate, blood, and BAL if beneficial) and daily decision-making on antibiotic requirements to de-escalate and avoid prolonged therapy that will lead to the development of antibiotic resistance.

The considerable variability in the incidence of secondary bacterial (4.8–42.8%) and fungal (0.9–33.3%) pulmonary infections reported and time taken for the diagnosis can be due to several limitations across the various observational studies included in this review. (1) The majority of the studies examining the incidence of secondary pulmonary infections are of poor quality and limited by the lack of a clear definition of secondary infections versus co-infections. There is also an absence of a standardized definition with the heterogeneity of diagnostic criteria used to differentiate between true invasive pulmonary fungal infection from colonization.[33–35] (2) Moreover, secondary pulmonary infections observed are a bystander (minor secondary outcome) result or identified during subgroup analysis while assessing the many characteristics, risk factors, and outcomes of hospitalized COVID-19 patients [3, 24, 30–32, 56, 58]. (3) It is not uncommon for early and late secondary infections to be frequently clustered together in the currently available literature for COVID-19 patients that may lead to the under- or overestimation of the exact incidence of secondary pulmonary infections, depending on the duration of the study period, especially among the 78.6% retrospective studies included in our review [67]. (4) The wide range of incidence rates reported for secondary pulmonary infections might have been due to the differences in the patient population, severity of illness, diagnostic sampling, and frequency of surveillance cultures obtained across various observational studies from multiple different countries. The routine use of bronchoscopy with BAL in critically ill COVID-19 patients where many are intubated and requiring IMV may lead to the over-diagnosis of secondary pulmonary infections [14–16, 43]. (5) The restricted search methodology that is confined to English literature as we (authors) are not well-versed in other languages during this global pandemic likely contribute to the under-recognition of the true incidence of secondary pulmonary infections. (6) Furthermore, the high mortality rate associated with COVID-19 pneumonia may be an independent competing risk factor for the development of

late secondary infection, leading to an unintended underestimation of the actual risk in non-deceased COVID-19 patients [67]. (7) Lastly, the widespread use of empirical antibiotics, analgesics, and corticosteroids likely mask underlying symptoms of infections, and lead to the delay and also underdiagnosis of secondary pulmonary infections. This could be due to the lack of routine surveillance cultures obtained because of fear towards COVID-19 transmission to health care professionals with prolonged patient contact [68]. These explain the variable incidence rate and inability to effectively perform a meta-analysis to determine better the incidence, risk factor, prognostic marker, and secondary pulmonary infection outcome in COVID-19 patients.

Conclusion

Our review on secondary pulmonary infections is limited by the lack of a clear definition of secondary infections versus co-infections, the inconsistency of the type microorganisms identified and time that surveillance cultures are obtained, the lack of information available on the associated antibiotic sensitivities of microorganisms, and duration of antibiotic treatment across various observational studies, small case reports and series, and variability in clinical characteristics reported in hospitalized COVID-19 patients. Additionally, with an observed strain being placed on the healthcare systems during the ongoing COVID-19 pandemic, there is a need for organized antimicrobial stewardship programs in the hospital to minimize the use of unnecessary empiric antibiotics and de-escalation of antibiotics when possible. As variation continues to exist on what constitutes a secondary infection (that we defined as infections occurring 48 h after admission) due to the lack of clear and consistent definition among many observational studies, we hope that a large, well-designed study can be performed in the future to accurately determine the incidence, microorganisms, risk factors, predictors, and outcomes of secondary pulmonary infections in hospitalized COVID-19 patients.

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Declarations

Conflict of interest None.

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